

# Package: macrosyntR (via r-universe)

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**Type** Package

**Title** Draw Ordered Oxford Grids

**Version** 0.3.4

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**Depends** R (>= 4.1.0)

**Imports** stats, utils, ggplot2, igraph, tidyr, reshape2, dplyr,  
stringr, rlang

**Description** Use standard genomics file format (BED) and a table of orthologs to illustrate synteny conservation at the genome-wide scale. Significantly conserved linkage groups are identified as described in Simakov et al. (2020) <doi:10.1038/s41559-020-1156-z> and displayed on an Oxford Grid (Edwards (1991) <doi:10.1111/j.1469-1809.1991.tb00394.x>) or a chord diagram as in Simakov et al. (2022) <doi:10.1126/sciadv.abi5884>. The package provides a function that uses a network-based greedy algorithm to find communities (Clauset et al. (2004) <doi:10.1103/PhysRevE.70.066111>) and so automatically order the chromosomes on the plot to improve interpretability.

**Encoding** UTF-8

**LazyData** true

**License** GPL-3

**URL** <https://github.com/SamiLh11/macrosyntR>

**BugReports** <https://github.com/SamiLh11/macrosyntR/issues>

**Roxygen** list(markdown = TRUE)

**RoxygenNote** 7.3.1

**Suggests** knitr, rmarkdown

**VignetteBuilder** knitr

**Repository** <https://samilh11.r-universe.dev>

**RemoteUrl** <https://github.com/samilh11/macrosyntR>

**RemoteRef** HEAD

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## Contents

compute_linkage_groups . . . . .	2
compute_macrosyteny . . . . .	3
get_syntenic_genes . . . . .	4
load_orthologs . . . . .	4
plot_chord_diagram . . . . .	6
plot_macrosyteny . . . . .	7
plot_oxford_grid . . . . .	8
reorder_macrosyteny . . . . .	10
reorder_multiple_macrosytenies . . . . .	11
reverse_species_order . . . . .	12
subset_linkage_orthologs . . . . .	13

**Index** **14**

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compute\_linkage\_groups  
*Compute Linkage groups*

---

### Description

This is a function to compute the conserved linkage groups shared between two or more species. It computes the significant associations between chromosomes of all species versus all (pairwise) using the fischer test implemented in compute\_macrosyteny(). It outputs a dataframe shaped as following : sp1.Chr,sp2.Chr,..., spN.chr,n,LGs where n is the number of shared orthologs in the group and LGs are the IDs for the linkage groups

### Usage

```
compute_linkage_groups(orthologs_df)
```

### Arguments

orthologs\_df    dataframe. orthologs with genomic coordinates loaded with load\_orthologs()

### Value

A dataframe object

## Examples

```
# basic usage of compute_linkage_groups:

orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")

my_orthologs <- read.table(orthologs_table, header=TRUE)

my_macrosynteny <- compute_linkage_groups(my_orthologs)
```

---

compute\_macrosynteny *Compute significant macrosynteny blocks*

---

## Description

This is a function to generate the contingency table of an orthologs dataframe and apply fisher test to calculate the significant associations. It outputs a dataframe shaped as following : sp1.Chr,sp2.Chr,a,pval,significant,pval\_a

## Usage

```
compute_macrosynteny(orthologs_df, pvalue_threshold = 0.001)
```

## Arguments

orthologs\_df      dataframe. orthologs with genomic coordinates loaded with load\_orthologs()  
pvalue\_threshold  
                  numeric. threshold for significancy. (default equals 0.001)

## Value

A dataframe object

## Examples

```
# basic usage of compute_macrosynteny :

orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")

my_orthologs <- read.table(orthologs_table, header=TRUE)

my_macrosynteny <- compute_macrosynteny(my_orthologs)
```

---

get\_syntenic\_genes      *get the syntenic genes as a table*

---

### Description

This is a function to extract all the syntenic genes from an orthologs\_df. It requires as input an orthologs\_df loaded by load\_orthologs().

### Usage

```
get_syntenic_genes(orthologs_df)
```

### Arguments

orthologs\_df      dataframe. orthologs with genomic coordinates loaded by load\_orthologs()

### Value

dataframe composed of details for each detected syntenic block of genes. It contains the following columns : sp1.Chr, sp1.Start, sp1.End, sp2.Chr, sp2.Start, sp2.End, size, sp1.IDs, sp2.IDs

### See Also

[load\\_orthologs\(\)](#)

### Examples

```
# basic usage of get_syntenic_genes :

orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")

my_orthologs <- read.table(orthologs_table, header=TRUE)

my_syntenic_block_of_genes <- get_syntenic_genes(my_orthologs)
```

---

load\_orthologs      *load orthologs with their genomic coordinates.*

---

### Description

Puts together the table of orthologous genes with their genomic coordinates in the two or more species. It outputs a data.frame shaped as following : sp1.ID,sp1.Chr,sp1.Start,sp1.End,sp1.Index,sp2.ID,sp2.Chr,sp2.Start,sp2.Index

**Usage**

```
load_orthologs(  
  orthologs_table,  
  sp1_bed = NULL,  
  sp2_bed = NULL,  
  bedfiles = NULL  
)
```

**Arguments**

orthologs_table	character. Full path to the orthologs table (format : geneID_on_species1 geneID_on_species2 geneID_on_speciesN)
sp1_bed	(deprecated) character. Full path to the genomic coordinates of the genes on species1
sp2_bed	(deprecated) character. Full path to the genomic coordinates of the genes on species2
bedfiles	array. List of full paths to the genomic coordinates ordered as in the appearing order of the orthologs_table (BED format)

**Value**

dataframe composed of genomic coordinates and relative index of orthologs on both species

**Examples**

```
# basic usage of load_orthologs for two species :  
  
orthologs_file <- system.file("extdata", "Bflo_vs_Pyes.tab", package="macrosyntR")  
bedfile_sp1 <- system.file("extdata", "Bflo.bed", package="macrosyntR")  
bedfile_sp2 <- system.file("extdata", "Pyes.bed", package="macrosyntR")  
  
my_orthologs <- load_orthologs(orthologs_table = orthologs_file,  
                              bedfiles = c(bedfile_sp1, bedfile_sp2))  
  
# example with 3 species :  
orthologs_file <- system.file("extdata", "Single_copy_orthologs.tsv", package="macrosyntR")  
bedfile_sp3 <- system.file("extdata", "Pech.bed", package="macrosyntR")  
  
my_orthologs <- load_orthologs(orthologs_table = orthologs_file,  
                              bedfiles = c(bedfile_sp1, bedfile_sp2, bedfile_sp3))
```

---

plot\_chord\_diagram     *plot the Macro-synteny as a chord diagram*

---

### Description

This is a function to plot the chord diagrams to compare the macro synteny of two or more species. It requires as input an orthologs\_df loaded by load\_orthologs()

### Usage

```
plot_chord_diagram(
  orthologs_df,
  species_labels = NULL,
  species_labels_size = 5,
  color_by = "sp1.Chr",
  custom_color_palette = NULL,
  reorder_chromosomes = TRUE,
  remove_non_linkage_orthologs = TRUE,
  species_labels_hpos = -400,
  label_size = 2,
  ideogram_fill = "white",
  ideogram_color = "black",
  ideogram_height = 4,
  gap_size = 40,
  ribbons_curvature = 0.1,
  ribbons_alpha = 0.5
)
```

### Arguments

orthologs\_df     dataframe. orthologs with genomic coordinates loaded by the load\_orthologs()

species\_labels   list of characters. names of the species to display on the plot

species\_labels\_size     integer. size of the labels (default = 2)

color\_by     string. name of the column in the orthologs\_df to color the links by (default = "sp1.Chr")

custom\_color\_palette     list of characters. palette to use for the coloring of the links following the argument color\_by

reorder\_chromosomes     logical. (default = TRUE) tells whether to reorder the chromosomes in clusters as implemented in reorder\_macrosynteny()

remove\_non\_linkage\_orthologs     logical. (default = TRUE) tells whether to remove the orthologs that are not within significant linkage groups as calculated by compute\_linkage\_groups().

species\_labels\_hpos  
(default = -400)

label\_size integer. size of the labels to display on the ideograms (default = 2)

ideogram\_fill character. name of the colors to fill the ideograms with (default = "white")

ideogram\_color character. name of the colors to draw the borders of the ideograms with (default = "black")

ideogram\_height  
integer. height of the ideograms (default = 4)

gap\_size integer. Size of the gap separating the ideograms (default = 40)

ribbons\_curvature  
float. curvature of the ribbons (default = 0.1)

ribbons\_alpha float. alpha of the ribbons (default = 0.5)

### Value

A ggplot2 object

### See Also

[load\\_orthologs\(\)](#)  
[reorder\\_macrosyteny\(\)](#)  
[compute\\_linkage\\_groups\(\)](#)

### Examples

```
# basic usage of plot_oxford_grid :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
plot_chord_diagram(my_orthologs, species_labels = c("B. flo", "P. ech"))
```

---

plot\_macrosyteny      *Plot Macro-syteny*

---

### Description

This is a function to generate the contingency table of an MBH dataframe and apply fisher test to calculate the significant associations.

### Usage

```
plot_macrosyteny(macrosynt_df, sp1_label = "", sp2_label = "")
```

**Arguments**

macrosynt_df	dataframe of contingency table with p-values calculated by the compute_macrosynteny() function
sp1_label	character. The name of the species1 to display on the plot
sp2_label	character. The name of the species2 to put on the plot

**Value**

ggplot2 object

**See Also**

[compute\\_macrosynteny\(\)](#)

**Examples**

```
# basic usage of plot_macrosynteny :
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")
my_orthologs <- read.table(orthologs_table, header=TRUE)
my_macrosynteny <- compute_macrosynteny(my_orthologs)

plot_macrosynteny(my_macrosynteny,
                  sp1_label = "B. floridae",
                  sp2_label = "P. yessoensis")
```

---

plot\_oxford\_grid      *plot the Macro-synteny as an oxford grid.*

---

**Description**

This is a function to plot the oxford grided plot to compare the macro synteny of two species. It requires as input an orthologs\_df loaded by load\_orthologs()

**Usage**

```
plot_oxford_grid(
  orthologs_df,
  sp1_label = "",
  sp2_label = "",
  dot_size = 0.5,
  dot_alpha = 0.4,
  reorder = FALSE,
  keep_only_significant = FALSE,
```



```

    color_by = NULL,
    pvalue_threshold = 0.001,
    color_palette = NULL,
    shade_non_significant = TRUE,
    reverse_species = FALSE,
    keep_sp1_raw_order = FALSE
  )

```

### Arguments

**orthologs\_df** dataframe. orthologs with genomic coordinates loaded by the `load_orthologs()`

**sp1\_label** character. name of 1st species to display on the plot

**sp2\_label** character. name of 2nd species to display on the plot

**dot\_size** numeric. (default = 0.5)

**dot\_alpha** numeric. (default = 0.4)

**reorder** logical. (default = FALSE) tells whether to reorder the chromosomes in clusters as implemented in `reorder_macrosynteny()`

**keep\_only\_significant**  
logical. (default = FALSE)

**color\_by** string/variable name. (default = NULL) column of the `orthologs_df` to use to color the dots.

**pvalue\_threshold**  
numeric. (default = 0.001)

**color\_palette** vector. (default = NULL) list of colors (as string under double quote) for the clusters. The amount of colors must match the amount of clusters.

**shade\_non\_significant**  
logical. (default = TRUE) When TRUE the orthologs located on non-significant linkage groups are displayed in "grey"

**reverse\_species**  
logical. (default = FALSE) When TRUE the x and y axis of the plot are reversed. sp1 is displayed on the y axis and sp2 is displayed on the x axis.

**keep\_sp1\_raw\_order**  
logical.(default equals FALSE) tells if the reordering should be constrained on the species1 and change just the order of the species2

### Value

A `ggplot2` object

### See Also

[load\\_orthologs\(\)](#)  
[reorder\\_macrosynteny\(\)](#)

**Examples**

```
# basic usage of plot_oxford_grid :

orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")

my_orthologs <- read.table(orthologs_table, header=TRUE)

plot_oxford_grid(my_orthologs,
                 sp1_label = "B. floridae",
                 sp2_label = "P. echinospica")

# plot a reordered Oxford Grid and color by cluster :

plot_oxford_grid(my_orthologs,
                 sp1_label = "B. floridae",
                 sp2_label = "P. echinospica",
                 reorder = TRUE,
                 color_by = "clust")
```

---

reorder\_macrosyteny    *Reorder the mbh\_df before plotting*

---

**Description**

This is a function to reorder an orthologs\_df, that was generated with load\_orthologs(). It retrieves communities using igraph::cluster\_fast\_greedy.

**Usage**

```
reorder_macrosyteny(
  orthologs_df,
  pvalue_threshold = 0.001,
  keep_only_significant = FALSE,
  keep_sp1_raw_order = FALSE
)
```

**Arguments**

orthologs\_df    dataframe. mutual best hits with genomic coordinates loaded with load\_orthologs()  
pvalue\_threshold    numeric. threshold for significancy. (default equals 0.001)  
keep\_only\_significant    logical. (default equals FALSE) tells if the non significant linkage groups should be removed. It drastically speeds up the computation when using one highly fragmented genome.  
keep\_sp1\_raw\_order    logical. (default equals FALSE) tells if the reordering should be constrained on the species1 and change just the order of the species2

**Value**

A dataframe object

**See Also**

[load\\_orthologs\(\)](#)

[compute\\_macrosynteny\(\)](#)

**Examples**

```
# basic usage of reorder_macrosynteny :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_orthologs_reordered <- reorder_macrosynteny(my_orthologs)
```

---

reorder\_multiple\_macrosynteny

*Reorder the chromosomes of two or more species before plotting*

---

**Description**

This is a function to reorder an orthologs\_df, same as reorder\_macrosynteny, but it handles tables with more than 2 species.

**Usage**

```
reorder_multiple_macrosynteny(orthologs_df)
```

**Arguments**

orthologs\_df     dataframe. orthologs with genomic coordinates loaded with load\_orthologs()

**Value**

A dataframe object

**See Also**

[load\\_orthologs\(\)](#)

[compute\\_macrosynteny\(\)](#)

[reorder\\_macrosynteny\(\)](#)

## Examples

```
# basic usage of reorder_macrosynteny :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_orthologs_reordered <- reorder_multiple_macrosyntenies(my_orthologs)
```

---

reverse\_species\_order *Reverse order of the species in an orthologs\_df.*

---

## Description

Returns an orthologs\_df (data.frame) with reversed species order compared to the inputted orthologs\_df. sp1 becomes sp2 and the otherway around. It intends at facilitating the integration of more than just two datasets. It outputs a data.frame shaped as following : sp1.ID,sp1.Chr,sp1.Start,sp1.End,sp1.Index,sp2.ID,

## Usage

```
reverse_species_order(orthologs_df)
```

## Arguments

orthologs\_df    orthologs\_df dataframe. mutual best hits with genomic coordinates loaded with load\_orthologs()

## Value

dataframe composed of genomic coordinates and relative index of orthologs on both species

## See Also

[load\\_orthologs\(\)](#)

## Examples

```
# basic usage of reverse_species_order :  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_orthologs_reversed <- reverse_species_order(my_orthologs)
```

---

`subset_linkage_orthologs`*Subset Orthologs contained in conserved linkage groups*

---

**Description**

This is a function to subset an `orthologs_df` and keep only the orthologs that are within significant linkage groups computed by the function `compute_linkage_groups()`.

**Usage**

```
subset_linkage_orthologs(orthologs_df, linkages = NULL)
```

**Arguments**

<code>orthologs_df</code>	dataframe. orthologs with genomic coordinates loaded with <code>load_orthologs()</code>
<code>linkages</code>	dataframe. table listing the linkage groups as returned by the function <code>compute_linkage_groups()</code>

**Value**

A dataframe object

**See Also**

[load\\_orthologs\(\)](#)

[compute\\_linkage\\_groups\(\)](#)

**Examples**

```
# basic usage of compute_linkage_groups:  
  
orthologs_table <- system.file("extdata", "my_orthologs.tab", package="macrosyntR")  
  
my_orthologs <- read.table(orthologs_table, header=TRUE)  
  
my_macrosynteny <- compute_linkage_groups(my_orthologs)
```

# Index

`compute_linkage_groups`, [2](#)  
`compute_linkage_groups()`, [7](#), [13](#)  
`compute_macrosynteny`, [3](#)  
`compute_macrosynteny()`, [8](#), [11](#)

`get_syntenic_genes`, [4](#)

`load_orthologs`, [4](#)  
`load_orthologs()`, [4](#), [7](#), [9](#), [11–13](#)

`plot_chord_diagram`, [6](#)  
`plot_macrosynteny`, [7](#)  
`plot_oxford_grid`, [8](#)

`reorder_macrosynteny`, [10](#)  
`reorder_macrosynteny()`, [7](#), [9](#), [11](#)  
`reorder_multiple_macrosynteny`, [11](#)  
`reverse_species_order`, [12](#)

`subset_linkage_orthologs`, [13](#)